

## In- Vitro Rooting and Hardening of Excised Shoots of *Momordica Cymbalaria* Hook. F.

S.V. Madhale., M. S. Sutare., S. P. Dorugade., V. M. Lagade and P. S. Pawar  
Department of Botany, Shri. Yashwantrao Patil Science College, Solankur, (Maharashtra), India.

---

### Abstract:

*In vitro* culture technique is now a days becoming a popular tool for the conservation of rare, endemic and endangered plants. In the family Cucurbitaceae *in-vitro* propagation of different members has been reported by several researchers. In the present research work *in-vitro* rooting and hardening is tried for excised shoots of *Momordica. cymbalaria*. Naphthalene Acetic Acid (NAA) and Indole Butyric Acid (IBA) were tried separately. IBA found more suitable for this purpose.

**Key Words:** *Momordica cymbalaria*, Rooting, Hardening, IBA, NAA

---

### ABBREVIATIONS:

IBA - Indole Butyric Acid, NAA - Naphthalene Acetic Acid, 2-4-Dichloro Phenoxy Acetic Acid, BAP- Benzyl Adenine Purine, MS-Murashige and Skoog's Media(1962).

### I. INTRODUCTION:

*Momordica cymbalaria* is a rare, wild and medicinally potential plant of cucurbitaceae family. It is endemic to certain parts of Maharashtra, Andhra Pradesh and Tamil Nadu states of India. The fruits are nutritious and utilized as vegetable (Chittapur, 2015). As the species is endemic and threatened, an attempt has been made by Madhale and Chavan (2009) to establish a protocol for its *in-vitro* culture. They have studied callus initiation and multishoot induction using Benzyl Adenine Purine (BAP) and Naphthalene Acetic Acid (NAA) as well as NAA and 2-4-Dichloro Phenoxy Acetic Acid (2-4-D) in Combination. Multishoot Induction is further studied for *in-vitro* Rooting and hardening. Therefore in the present piece of work an attempt has been made to study using NAA and Indole Butyric Acid (IBA) separately.

### II. MATERIAL AND METHODS:

The plant material of *M. cymbalaria* is collected from the different habitats Solapur dist. of Maharashtra and maintained in the botanical garden of Shivaji University, Kolhapur. Multishoot induction was achieved by following protocol standardized by Madhale and Chavan (2018). Further, excised shoots were cultured on half strength of MS medium containing NAA and IBA(0.1 to 4.0 mg /liter) along with 0.5 % activated charcoal for rooting. Hardening is studied as described by Gamborg and Philips (2004). For the process of hardening plastic cups ( 5cm diameter and 6 cm height ) were filled with hardening mixture (soil + sand + compost in 1:1:1 proportions) were used. The rooted shoots were carefully removed from tubes and transplanted in hardening mixture. These cups were kept in culture room for two weeks to avoid shock due to environmental changes and desiccation. After that these cups were transferred to shade net with 50% light cut off. Finally plantlets were transferred to field.

### III. RESULTS AND DISCUSSION:

The growth regulator NAA and IBA were used separately in the concentration of 0.2 to 0.3 mg/l to study rooting in excised shoots. Positive response of rooting started in the concentration of 0.5 mg/lit. and 1.0 mg /liter in case of NAA and IBA respectively. In case of NAA and IBA positive response gradually increased up to 2.0 mg/liter concentration and started lowering. The response is three fold in IBA than NAA. (Mahipal *et al.*, 2016 and 2017).

The growth regulator IBA was found to be most effective in root induction. (Mahipal *et al.*, 2016). At low concentration of IBA shows 8% rooting induction. The gradual increase in IBA concentration increases the percentage of rooting in culture. IBA 2.0mg/l was found to be most effective for rooting. (M. Chabukswar and Deodhar, 2005) More than 2.0 mg/l concentration of IBA gradually decreases the percent of rooting. (Table No.1) The IBA exhibited best results than the NAA. (Phlomina and Rao, 1999; Segio *et al.*, 2000; Johnson, 2002; Vasanth *et al.*, 2002; Biljana, *et al.*, 2004).

Root induction in case of IBA started after 4 days of culture. It continues almost up to 15 days. Later on the rooted shoots were transplanted in hardening mixture. The plantlets in hardening mixture were kept for 5 days under laboratory conditions. The hardening process took 20 days. The plantlets were hardened previously under laboratory conditions and then under 50% shed net at 80% humidity and 25-30°C temperature. The hardened plantlets were then transferred to the field for further establishment.

The success rate of hardening process is 80%. No adverse morphological effects were observed except slight wilting. Root: shoot ratio is also depicted in the table. Interestingly it is high in case of 2.5 mg/liter IBA, where root induction is comparatively less. Therefore, 2.0 mg/liter IBA is the best for root induction and growth.

**Table No.-1:** Effect of NAA and IBA on rooting of excised shoots of *M. cymbalaria*

Growth regulator in mg/l	% cultures showing response			
	NAA		IBA	
	% of root induction	Ratio of root/shoot	% of root induction	Ratio of root/shoot
0.2	.....	.....	.....	.....
0.5	8.0 ±0.0	1.0±0.0	.....	.....
1.0	14.5±1.0	2.0±0.0	08±0.0	3±0.0
1.5	10.0±0.0	2.0±0.2	18±0.0	5±0.8
2.0	18.0±2.5	3.0±0.0	68±2.5	10±2.2
2.5	9.0±0.0	1.0±0.0	23.2±1.9	11±2.0
3.0	2.0±0.6	1.0±0.2	10.0±0.0	8±0.0

(culture period 4 weeks)



**In- vitro rooting and hardening of excised shoots of *M. cymbalaria***

- A) Profusely root initiation
- B) Rooting initiation - I stage
- C) Hardening of excised shoots

**IV. CONCLUSION:**

The above experimental results indicate that, IBA is superior than NAA in root induction. The significance of IBA in root induction has been reported in *Cucumis sativus* by Shirgave and Chavan, 2001; Misra and Bhatnagar, 1995. The success rate of hardening process is 80%. Overall sixty five days are required to establish new plantlet in the field. Among this period thirty days are required for multishoot development (time up to the excision). 15 days require rooting initiation and 20 days require to hardening process.

**REFERENCES:**

- [1]. Biljana, B., Dunja, L., Levanic, Snjezana, M., Srecko J., and Sibila, J. (2004). Formation of embryogenic callus in hairy roots of pumpkin (*Cucurbita pepo* L.); *In Vitro Cell. Dev. Biol. Plant.* 40:182–187.
- [2]. Chittapur R. (2015) *Momordica cymbalaria* a nutritious underutilized vegetable taxonomy, nutritional, medicinal, propagation, hybridization and cytological aspects, *International Journal of Agricultural Science and Research* ,Vol. 5, Issue 4, 255-262
- [3]. Gamborg, O.L; and Phillips, G.C. (2004). Plant Cell, Tissue and Organ culture Fundamental methods, Narosa publishing House, New Delhi, India, 420.
- [4]. Johnson, M., Villinayagam, S., Manickam, V. S. and Seeni, S. (2002). Micropropagation of *Rhinacanthus nasutus* (L.) Kurz. A medicinally important plant. *Phytomorphology*. 52(4): 331-336.
- [5]. Madhale, S. V. and N. S. Chavan (2009). Ecological status and identification of threat level of *Momordica cymbalaria* Hook. f. from some parts of Maharashtra and Karnataka. *Int j. for Usuf. mngt.* 10(2): 66-70.
- [6]. Madhale, S. V. and N. S. Chavan (2018). *In-vitro* conservation of *Momordica cymbalaria* Hook. f. by Leaf culture. *International journal of trends in scientific and research development*. Vol. 2(3), 882.
- [7]. Mahipal S. Shekhawat, N. Kannan M. Manokari, C.P. Ravindran. (2015). *In vitro* regeneration of shoots and ex vitro rooting of an important medicinal plant *Passiflora foetida* L. through nodal segment cultures, *Journal of Genetic Engineering and Biotechnology*, Volume 13, 2, 209-214. <https://doi.org/10.1016/j.jgeb.2015.08.002>
- [8]. Mahipal S. Shekhawat & M. Manokari (2016). In vitro regeneration frequency, micro- morphological studies and ex vitro rooting of *Hemidesmus indicus* (L.) R. Br.: a multi- potent endangered climber. *Indian Journal of Plant Physiology* ,volume 21, pages151–160
- [9]. Mahipal S. Shekhawat, M. Manokari & J. Revathi (2017) *In vitro* propagation and *ex vitro* rooting of *Aerva lanata* (L.) Juss. ex Schult.: a rare medicinal plant. *Indian Journal of Plant Physiology*. Vol. 22, pages40–47
- [10]. Mahipal S. Shekhawat and M. Manokari. (2015) Efficient *In Vitro* Propagation by *Ex Vitro* Rooting Methods of *Artemisia absinthium* L., an Ethnobotanically Important Plant. *Chinese Journal of Biology*, Article ID 273405 | <https://doi.org/10.1155/2015/273405>
- [11]. Meera M Chabukswar and Manjushri Deodhar (2005) Rooting and hardening of in vitro plantlets of *Garcinia indica* Chois. *Indian Journal of Biotechnology* 4(3)
- [12]. Misra, A. K. and Bhatnagar, S. P. (1995). Direct shoot regeneration from the leaf explant of cucumber (*Cucumis sativus* L.) *Phytomorphology* 45 (1 and 2); 47-55.
- [13]. Phlomin, N. S. and Rao, J. V. S. (1999). Multiple shoot regeneration from seed cultures of soapnut (*Sapindus mukurossi* Gaertn.) *Phytomorphology* 49(4): 419-423.
- [14]. Segio, E. Simone, B. Fernando, F. and Lucian, A. (2000). Clonal propagation of Roman chamomile (*Anthermis nobilis* L. ) *J. Herbs Species and medicinal plants*. 7 : 35-40.
- [15]. Shirgave P. D. and Chavan, N. S. (2001). *In vitro* culture of *Momordica dioica* Roxb. Ex. Wild. Asian Jr. of Microbial. Biotech. And Env. Sc.:3(3): 173-175.
- [16]. Vasanth, K., Lakshmi Prabha, A., Jayakumar, M., Mathuswamy, A. and Jayabalan, N. (2002). *In vitro* plant regeneration from shoot tip explant of *Panicum sumatrense*. *Pt. Cell Biot. And Mol. Biol.* 3 (3 and 4) : 111-116.